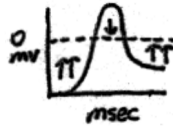


Lecture notes courtesy of Wyan-Ching Mimi Lee. Used with permission.

3/3/04

muscle AP



reversal potential = 0 (or -15 mV, depending on muscle cell)

b/c AChR lets in  $\text{Na}^+$  and  $\text{K}^+$

- collagenase to get postsynaptic side clear, patch clamp (w/ ACh in blunt microelectrode), look at current flow (magnitude + direction)

↳ voltage clamp

( $V_m < 0$ , current negative = inward)

$V_m > 0$ , current positive = outward)

- ionotropic receptors have reversal potentials

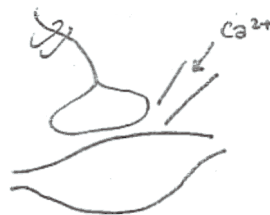
- if  $E_{\text{reversal}}$  above or below threshold, determines if excitatory or inhibitory

- if  $E_{\text{reversal}} = \text{threshold}$ , will not affect either way

- stimulate extracellularly presynaptic, record intracellularly postsynaptic

- need  $\text{Ca}^{2+}$  (4) for presynaptic transmitter release, EPSP

- cobalt interferes w/ natural  $\text{Ca}^{2+}$  leak from slightly damaged muscle



$\text{Ca}^{2+}$  must be near synaptic cleft, present around when AP reaches terminal (or right before reaches terminal)

treated whole system w/ TTX, increased stimulus intensity to passively conduct to terminal:

found that pure voltage could mimic AP: evidence for voltage-gated  $\text{Ca}^{2+}$  channels

- intracellular presynaptic injection of  $\text{Ca}^{2+}$  also gives EPSP

- minis - correspond to small releases of ACh (b/c can affect w/ curare, neostigmine)

- make up mEPSP: reduce  $\text{Ca}^{2+}$ , stimulate presynaptically, record postsynaptically

- effect comes in quantal sizes (Mn)

- Gaussian distribution around 0.4 mV

- ↳ direct evidence for quantal transmission

- large number of vesicles w/ equal & independent probability of release (from Poisson curve assumptions)

Poisson curve assumptions)

- make preparations of synapses, synaptic vesicles
  - synaptobrevin - V-SNARE (important in exocytosis)
    - target of botulinum toxin (BOTOX) & tetanus toxin
    - mutants homologous to yeast mutants w/ exocytotic defects
      - yeast cycle vesicles between inner & outer lamellae
      - bad-cycling due to mutant v-snares & t-snares
      - vesicle endocytosis looked at in cell-free dog pancreas system; these same proteins showed up
  - synaptotagmin -  $Ca^{2+}$  binding domain: this is the calcium sensor for vesicle exocytosis

- neuromuscular junctions can be modulated
  - (eg by chip screaming - potentiated by adrenaline)
  - potentiated by sympathetic stimulation (neuromuscular transmission)
    - Orbelli effect (controversy: presynaptic or postsynaptic?): turns out is both
      - do quantal analysis before & after sympathetic stimulation: more quanta released per action potential will give more double, triple, etc releases (average # releases), if presynaptic effect; if postsynaptic (eg more ACh Rs, or more sensitive ACh Rs, will not affect placement of peaks by shifting histogram right (eg 0.4 mV  $\rightarrow$  0.6 mV)

-  $P(x) = e^{-m} \frac{m^x}{x!}$  (but there is better way)

So: if presynaptic  $\rightarrow$  easy to tell 0's from 1's :  $P_0 = e^{-m} \left( \frac{m^0}{0!} = 1 \right)$   
 if postsynaptic  $\rightarrow$  (increase mean quantal content) ( $P_0$  decreases)  
 if postsynaptic, will affect quantal size  
 measure fraction of failures, see if goes up or down

- mean quantal content = average # quanta (vesicles) released per stimulus (not contents of vesicle)
  - measure fraction of failures, plug into  $P_0 = e^{-m}$  (no dimensions)
    - $\rightarrow$  fraction of failures
    - higher quanta release # = lower probability of failure

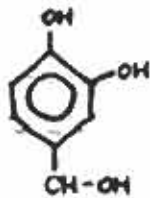
to measure change in quantal size, look at minis, look at size of mEPSPs

- this will increase if Orbelli effect postsynaptic

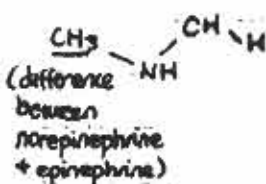
- quantal size  $\bar{V}_1 =$  average voltage displacement of quantum of spontaneous miniature potential  
not physical size of vesicle (all ~ same size, same density of transmitter)

Orbelli effect: <sup>norepinephrine</sup> more adrenergic comes down sympathetic axons to neuromuscular junctions

- adrenal glands secrete another compound (cycles around your body): <sup>norepinephrine?</sup> epinephrine
- these result in presynaptic enhanced quantal release. (NE)
- also postsynaptic response from circulating hormone (EPI)



phenylethylamine → tyrosine → tyramine → dopamine → noradrenaline  
 adrenaline ← (norepinephrine) → NE



adrenaline from adrenal medulla (on top of kidney)  
 - epinephrine same in Greek  
 ↳ EPI

norepinephrine + epinephrine both from phenylethylamine, differ by methyl group

antagonist: <sup>β-propranolol</sup> isoproterenol (EPI antagonist) <sup>heart conditions</sup> drug used for asthma in hospitals, stage fright  
 - applying this gives no change in quantal size  
 - better music performance

epinephrine - agonist = isoproterenol  
 (adrenaline) antagonist = β-propranolol (block adrenergic response on postsynaptic side)

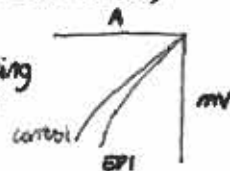
presynaptic = look at number of failures (Cuba found less failures)

norepinephrine - agonist = norepinephrine  
 (noradrenaline) antagonist = clonidine (blocks presynaptic response, gives pure postsynaptic)

postsynaptic = look at quantal size (will be bigger w/ Orbelli-effect)

- postsynaptic so increase actually from epinephrine closing leakage channels

depolarizing current leaks out less if RT ↑ (R input)  
 $V = IR$   
 so RT, VT depolarizes more + longer



I from ionotropic AChRs: doesn't change, depolarizes cell, but leaks out b/c muscle is big; plug up leakage holes in muscle, increase R

- modulated synapses - have volume control

- norepinephrine + epinephrine are modulatory transmitters (gain control)

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